PLANT METABOLITES. TRITERPENOID SAPONINS FROM CALENDULA ARVENSIS

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Calendula arvensis L. (Compositae) is an herbaceous plant used in Italian folk medicine as an antiinflammatory and antipyretic. In a recent pharmacologic study, the methanolic extract of aerial parts showed antiinflammatory activity (1). In previous work we reported the isolation and structure determination of a sesquiterpene glycoside from *C. arvensis* (2). In the present paper, we report the isolation of four triterpenoid saponins [1-4].

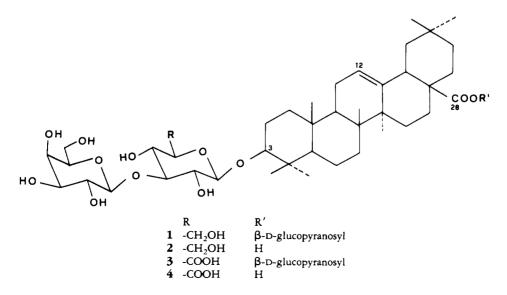
The aerial parts of air-dried plant material were extracted successively with light petroleum ether, $CHCl_3$ and MeOH. The MeOH extract was chromatographed on Sephadex LH-20, and the saponin-containing fractions were further purified by droplet counter current chromatography (dccc) (10) and hplc to obtain saponins **1-4**.

The molecular formulae $C_{48}H_{78}O_{18}$, $C_{42}H_{68}O_{13}$, $C_{48}H_{76}O_{19}$, $C_{42}H_{66}O_{14}$, for saponins **1-4**, respectively, were determined by DEPT ¹³C nmr (Table 1) and fabms, negative ion mode, in a thioglycerol-glycerol matrix (see Experimental).

Acid methanolysis liberated methyl galactoside and methyl glucoside (molar ratio 1:2) from 1, methyl galactoside and methyl glucoside (1:1) from 2, methyl galactoside, methyl glucuronide, and methyl glucoside (1:1:1) from 3, and methyl glucuronide and methyl glucuronide and methyl galactoside (1:1) from 4. The methylated sugars were analyzed by glc.

Signals in fabms of **3** and **4** at m/z 631 and 455, corresponded to the subsequent loss of a galactosyl moiety and a glucuronic moiety and indicated that glucuronic acid was attached to the aglycone. The aglycone (molecular formula C₃₀H₄₈O₃, deduced from fabms) for saponins **1**-**4**, was identified as oleanolic acid on the basis of ¹H-, ¹³Cand DEPT ¹³C-nmr spectral data and comparison with literature data (3,4).

The β -D-pyranosyl configuration of galactose, glucose, and glucuronic acid in saponins 1-4 was deduced from ¹H-and ¹³C-nmr spectra (see Table 1 and



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| Carbon | | Compounds | | | | DEPT |
|--------------------|----------------------------------|---|---|--|--|--|
| | | 1 | 2 | 3 | 4 | |
| Aglycone | 1 2 3 4 5 | 39.9 27.0 90.8 40.1 57.1 | 39.9 27.0 90.8 40.2 57.2 | 39.9 26.9 90.6 40.2 57.1 | 40.0 27.1 90.7 40.1 57.1 | CH ₂ CH ₂ CH C C C C |
| | 6 7 8 9 | 19.3 33.2 40.8 48.1 ^a | 19.4 33.5 40.9 48.1 ^a | 19.4 33.2 40.8 48.1 ^a | 19.4 33.2 40.9 48.1* | CH ₂ CH ₂ C C CH |
| | 10 11 12 13 14 | 37.9 24.6 123.8 144.8 43.0 | 38.0 24.6 123.6 145.4 43.1 | 37.9 24.6 123.9 144.8 43.0 | 37.9 24.6 123.9 145.3 43.1 | C CH ₂ CH C C |
| | 15 16 17 18 | 28.9 24.0 48.1 ^a 42.6 | 29.0 24.3 48.1 ^a 42.9 | 28.9 24.1 48.1ª 42.7 | 28.9 24.2 48.1 ^a 42.9 | CH ₂ CH ₂ C CH |
| | 19 20 21 22 23 | 47.3 31.5 34.9 34.1 28.7 | 47.5 31.6 35.1 34.2 28.6 | 47.3 31.5 35.0 34.1 28.6 | 47.4 31.6 35.0 34.1 28.5 | CH ₂ C CH ₂ CH ₂ CH ₂ CH ₃ |
| | 24 25 26 27 28 29 | 17.1 16.0 17.8 26.4 178.1 33.5 | 17.0 15.9 17.9 26.4 182.2 33.5 | 17.0 16.0 17.9 26.3 178.2 33.4 | 17.0 16.0 17.9 26.3 182.2 33.4 | CH ₃ CH ₃ CH ₃ CH ₃ C CH ₃ C CH ₃ |
| Glucose I | 30 1 2 3 4 5 | 24.0 95.7 73.9 78.5 71.3 78.4 | 24.0 | 24.0 95.8 74.1 78.5 71.4 78.4 | 24.0 | CH CH CH CH CH CH CH |
| Glucose II | 6 1 2 3 4 5 | 62.8 105.7 74.8 88.2 70.4 77.2 | 105.8 74.8 88.3 70.4 77.3 | 62.7 | | CH₂ CH CH CH CH CH |
| Glucuronic Acid | 6 1 2 3 4 5 | 62.9 | 63.0 | 105.2 74.9 86.2 72.3 77.2 | 105.3 74.8 86.2 72.2 77.2 | CH₂ CH CH CH CH CH |
| Galactose | 6 1 2 3 4 5 | 106.2 73.1 75.0 70.4 77.1 | 106.2 73.1 75.0 70.4 77.1 | 176.4 106.3 72.8 75.1 70.6 77.1 | 176.3 106.2 72.9 75.0 70.5 77.0 | C CH CH CH CH CH |
| | 6 | 62.8 | 62.6 | 62.7 | 62.7 | CH ₂ |

TABLE 1. ¹³C-nmr Data for Saponins 1-4 in CD₃OD

^aUnder CD₃OD signal.

Experimental section). The anomeric proton signal at δ 5.41 (1H, d, J=7.5 Hz) in the ¹H-nmr spectra and the anomeric carbon signals at δ 95.7 and 95.8 in the ¹³C-spectra of **1** and **3**, respectively, were attributed to a β -glucose unit linked to the 28-carboxyl group of the aglycone through an ester bond.

Basic hydrolysis of 1 gave saponin 2, while basic hydrolysis of 3 gave 4.

In saponins 2 and 4 the free carboxyl group of the aglycone appeared at δ 182.2 (δ 179.8 in pyridine- d_5 for 2), whereas, when esterified with the glucosyl moiety in saponins 1 and 3, it resonated at δ 178.1 (δ 176.2 in pyridine- d_5 for 1) (5,6). The C-3 of the aglycone as glycosidation site and the interglycosidic linkages in saponins 1-4 were derived from ¹³C-nmr data (Table 1). The ¹³C-nmr sugar signals of 1 and 2 indicated that the β -D-galactopyranosyl is the terminal unit and that this sugar is attached at the position 3 of the inner glucosyl unit (4,7,8).

Thus, the structure of saponin 1 is established as 3β -O-[β -D-galactopyranosyl-(1-3)- β -D-glucopyranosyl] oleanolic acid-28-O- β -D-glucopyranoside and that of 2 as 3β -O-[β -D-galactopyranosyl-(1-3)- β -D-glucopyranosyl] oleanolic acid. Saponins 1 and 2 are new natural compounds.

In saponins **3** and **4** the terminal sugar is the β -D-galactopyranosyl, attached at the position 3 of the β -Dglucopyranosyluronic acid (Table 1) (6). The structure of saponin **3** is established as 3β -O-[β -D-galactopyranosyl-(1-3)- β -D-glucopyranosyluronic acid] oleanolic acid-28-O- β -D-glucopyranoside, and that of **4** as 3β -O-[β -D-galactopyranosyl-(1-3)- β -D-glucopyranosyluronic acid] oleanolic acid.

Saponins 3 and 4 were earlier reported from *Calendula officinalis* L. (9). However, the present communication further confirms the assigned structure from spectral information and from hydrolytic studies.

EXPERIMENTAL

INSTRUMENTAL.-¹H- and ¹³C-nmr spectra were recorded on a Bruker WM-250 spectrometer. Chemical shifts are reported relative to TMS. The DEPT experiments were made by using polarization transfer pulse of 90° and 135°, respectively, obtaining in the first case only CH groups and in the other case positive signals for CH and CH₃ and negative ones for CH₂ groups. Polarization transfer delays were adjusted to an average CH coupling of 135 Hz. Fab-mass spectra were recorded on a Kratos MS-50 mass spectrometer equipped with a Kratos fab source. The spectra were obtained by dissolving the samples in a thioglycerol-glycerol matrix and placing them on a copper probe tip prior to bombardment with Ar atoms of energy 2-6 Kv. Optical rotations were recorded on a Perkin-Elmer 241 polarimeter. Dccc separations were performed on a DCC-A apparatus manufactured by Tokyo Rikakikai Co., equipped with 300 tubes. Hplc separations were performed on a Waters Model 6000A pump equipped with a U6K injector and differential refractometer Model 401 detector. Glc analyses were performed with a Carlo Erba Fractovap 2900 capillary column.

ACIDIC METHANOLYSIS OF 1-4, SUGAR ANALYSIS.—A solution of each saponin (0.5, 1 mg) in anhydrous 2 M HCl-MeOH (0.1 ml) was heated at 80° in a stoppered reaction vial for 10 h. After being cooled, the reaction mixture was neutralized with Ag_2CO_3 and centrifuged; the supernatant was evaporated to dryness. The residue was dissolved in TRISIL Z [0.05 ml; N-(trimethylsilyl) imidazole in pyridine, Pierce Chemical Co.], left at room temperature for 15 min, and analyzed by glc (25 m SE-30 capillary column, 146°). Glc peaks in the silylated hydrolysate co-eluted with those in silylated standards.

BASIC HYDROLYSIS.—The saponins 1 (20 mg) and 3 (10 mg) in 0.5 M aq. KOH (1 ml) were heated at 110° in a stoppered reaction vial for 2 h. The reaction mixture was adjusted to pH 7 and then extracted with BuOH. The organic phase was evaporated to dryness, dissolved in CD₃OD, and analyzed by ¹H nmr and ¹³C nmr.

EXTRACTION AND ISOLATION.—Plants of *C.* arvensis were collected near Naples, Italy, in the Spring 1985; a sample has been deposited in Dipartimento di Chimica delle Sostanze Naturali, University of Naples. The aerial parts of the airdried plant material (800 g) were extracted successively with light petroleum ether (40° - 70° bp) (8.6 g), CHCl₃ (7.5 g), and with MeOH (15 g); 2.2 g of the MeOH extract was chromatographed on Sephadex LH-20 (3×60 cm; MeOH; 8-ml fractions were collected) to yield 900 mg (fractions 39-43) of the saponin mixture. Purification was continued by dccc with CHCl₃-MeOH-H₂O (7:13:8) in which the stationary phase consisted of lower phase; ascending mode; flow 12 ml/h; 4-ml fractions were collected to yield a mixture I containing the more polar glycosides **1** and **3** (300 mg) in the fractions 52-63 and a mixture II containing **2** and **4** (200 mg) in the fractions 71-83. Both mixtures I and II were submitted to hplc on a C₁₈ μ -bondapak column (30 cm×7.8 mm i.d.) with MeOH-H₂O (65:35), to yield pure **1** (98 mg), **2** (30 mg), **3** (15 mg), and **4** (10 mg).

3-0-[β -D-GALACTOPYRANOSYL-(1-3)- β -D-GLU-COPYRANOSYL] OLEANOLIC ACID-28-0- β -D-GLU-COPYRANOSIDE [1].—Hplc retention time 12 min; [α]D=5° (MeOH); fabms, negative ions, glycerol-thioglycerol matrix, m/z 941 {(M-H)⁻], 779 ([(M-H)-162]⁻), 617 ([(M-H)-(162×2)]⁻), 455 ([(M-H)-(162×3)]⁻); ¹³C nmr in CD₃OD see Table 1; ¹H nmr in CD₃OD (aglycone) singlets at δ 0.833, 0.874, 0.949, 0.968, 0.988, 1.095, 1.195 (each 3H), δ 5.28 (1H, m, H-12); (sugars) δ 4.41 (1H, d, J=7.5 Hz, H-1 of galactose unit), δ 4.58 (1H, d, J=7.6 Hz, H-1 of glucose II unit), δ 5.41 (1H, d, J=7.5 Hz, H-1 of glusinglets I unit).

¹³C-nmr in pyridine- d_5 δ's of the aglycone corresponded to those of oleanolic acid (4); sugar signals: glucose I 95.6 (C-1), 73.9 (C-2), 78.7 (C-3), 71.2 (C-4), 78.9 (C-5), 62.1 (C-6); glucose II 105.9 (C-1), 74.8 (C-2), 89.0 (C-3), 70.0 (C-4), 77.5 (C-5), 62.6 (C-6); galactose 106.1 (C-1), 72.8 (C-2), 74.9 (C-3), 69.8 (C-4), 77.1 (C-5), 62.3 (C-6).

Basic hydrolysis of 1 afforded the saponin 2 (comparison of ¹H- and ¹³C-nmr spectra in CD₃OD). Acidic methanolysis of 1 afforded methyl glucoside ($\times 2$) and methyl glactoside.

3β-0-[β-D-GALACTOPYRANOSYL-(1-3)-β-D-GLU-COPYRANOSYL] OLEANOLIC ACID [2].—Hplc retention time 47 min, [α]D=8° (MeOH); fabms, negative ions, 779 ([M-H]⁻), 617 ([(M-H)-162]⁻), 455 ([(M-H)-(162×2)]⁻); ¹³C nmr in CD₃OD see Table 1; ¹H nmr in CD₃OD (aglycone) singlets at δ 0.852, 0.873, 0.936, 0.973, 0.986, 1.09, 1.19 (each 3H), δ 5.28 (1H, m, H-12); (sugars) δ 4.41 (1H, d, J=7.5 Hz, H-1 of glactose unit), δ 4.58 (1H, d, J=7.6 Hz, H-1 of glucose unit). ¹³C nmr in pyridine-d₅ δ's of the aglycone corresponded to those of oleanolic acid (4); sugar signals: see δ's values reported for glucose II and galactose sugar units of **1** (±0.2 ppm).

3β-0-[β-D-GALACTOPYRANOSYL-(1-3)-β-D-GLU-COPYRANOSYLURONIC ACID] OLEANOLIC ACID-28-0-β-D-GLUCOPYRANOSIDE [**3**].—Hplc retention time 9 min, $\{\alpha\}D=15^{\circ}$ (MeOH); fabms, negative ions, 955 ([M-H]⁻), 793 ([(M-H)-162]⁻), 631 ([(M-H)-(162×2)]⁻), 455 ([(M-H)-(324+176)]⁻); ¹³C nmr in CD₃OD see Table 1; ¹H nmr in CD₃OD (aglycone) singlets at δ 0.827, 0.86, 0.943, 0.962, 0.974, 1.072, 1.184 (each 3H), δ 5.28 (1H, m, H-12); (sugars) δ 4.41 (1H, d, J=7.5 Hz, H-1 of galactose unit), δ 4.61 (1H, d, J= 7.6 Hz, H-1 of glucuronic acid unit), δ 5.41 (1H, d, J=7.5 Hz, H-1 of glucose unit).

Basic hydrolysis of saponin 3 afforded saponin 4 (comparison of 1 H-nmr spectra in CD₃OD). Acidic methanolysis of 3 afforded methyl glucoside, methyl galactoside, and methyl glucuronide.

3β-0-[β-D-GALACTOPYRANOSYL-(1-3)-β-D-GLU-COPYRANOSYLURONIC ACID) OLEANOLIC ACID [4].—Hplc retention time 43 min, [α]D=12° (MeOH); fabms, negative ions, m/z 973 ([(M-H)⁻]), m/z 631 ([(M-H)-162]⁻), 455 ([(M-H)-(162+176)]⁻); ¹³C nmr in CD₃OD see Table 1; ¹H nmr in CD₃OD (aglycone) singlets at δ 0.852, 0.873, 0.936, 0.973, 0.986, 1.072, 1.187 (each 3H), δ 5.28 (1H, m, H-12); (sugars) δ 4.41 (1H, d, J=7.5, H-1 of galactose unit), δ 4.60 (1H, d, J=7.6 Hz, H-1 of glucuronic acid unit).

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